

Lecture 1: Cancer Biology for Modellers

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Motivation

- To be effective biomathematicians, we need to “speak biology” as well mathematics and programming.
 - Helps in communicating with team members
 - Biological understanding helps inform:
 - Model formulation and analysis
 - Evaluation of modelling predictions
 - Parameter estimation
 - Calibration
 - relates to image processing
 - Need to understand what you’re seeing (histopathology)

Lecture Outline

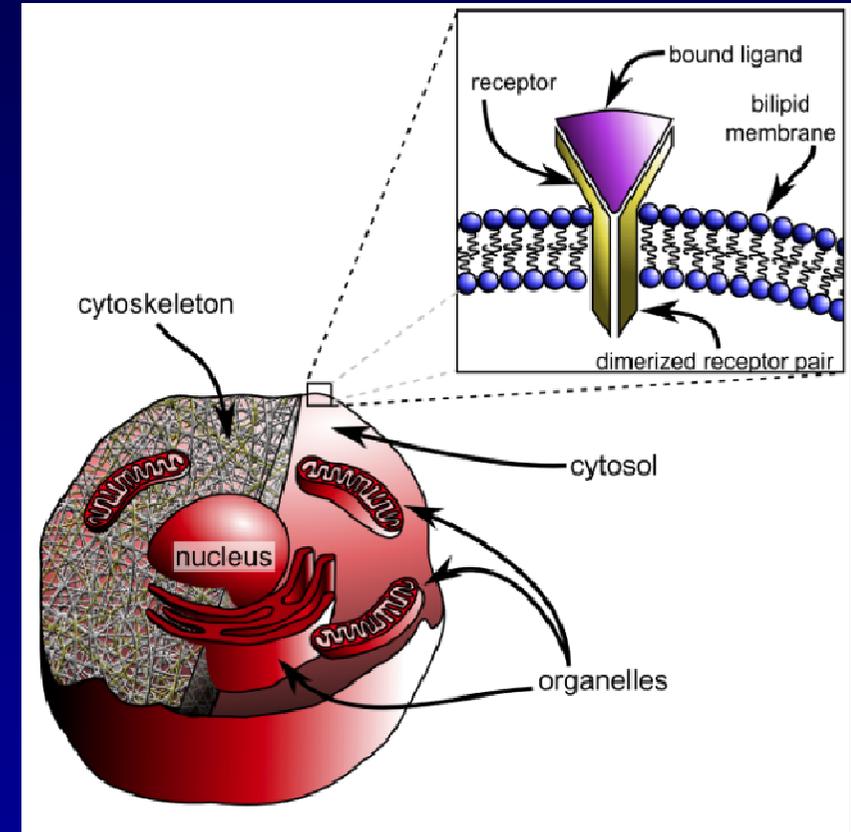
- Cell organisation
- Tissue organisation
- Maintaining tissue structure
- Cell birth and death
- Cell signalling
- Cell motility
- Oncogenes and tumour suppressor genes
- Abusing the system: cancer progression
- Coming next
- References and resources

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Cell Organisation

- Bilipid layer surrounding cytoplasm
 - Permeable to small molecules
 - Requires active transport of others
 - Ion pumps control cell volume, pH, etc.
 - Impermeable to larger proteins
- Cytoskeleton provides structure
- Organelles embedded in the cytoplasm to perform specialised functions
- Relatively rigid nucleus in the centre
- Receptors on cell surface:
 - Mediate cell-microenvironment communication by binding to ligands
 - Mechanically link cell cytoskeleton to membrane and microenvironment

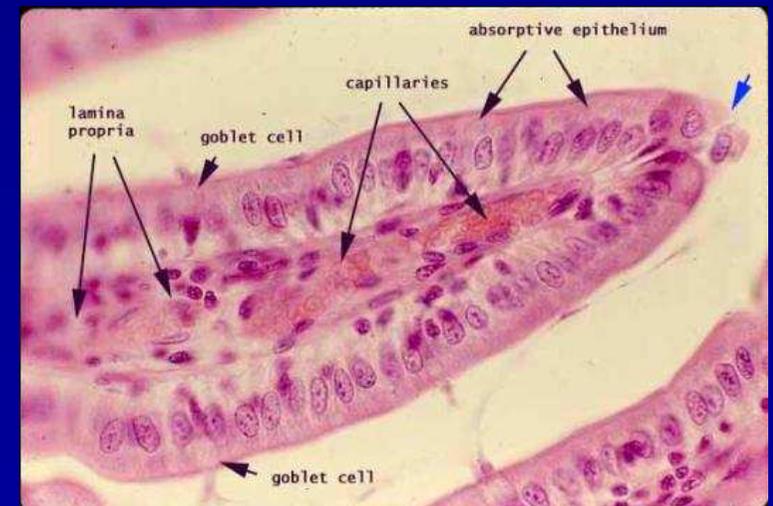
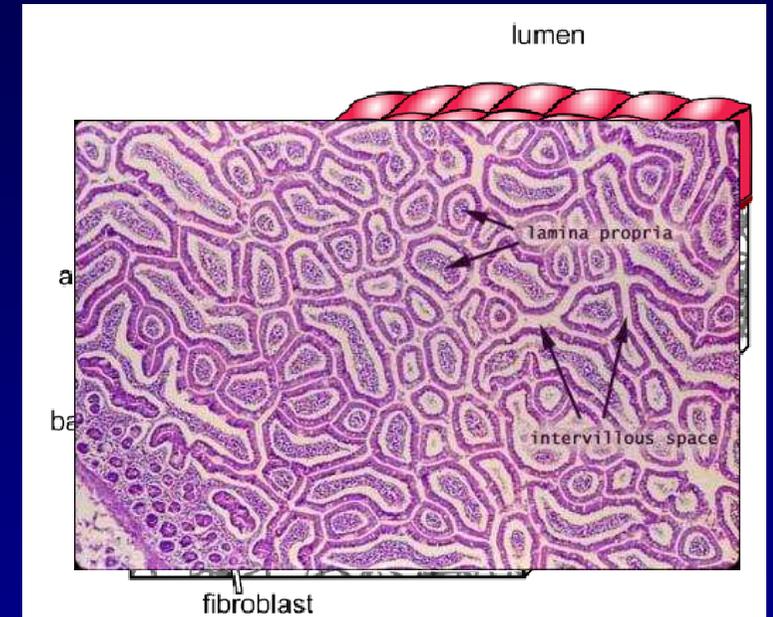


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Tissue Organisation

- **Typical tissue microstructure in an organ:**
 - **Epithelial tissue (epithelium)**
 - Sheets of specialised cells that perform the work of the organ (secretory products, filtration, etc.)
 - **Loose connective tissue (stroma)**
 - Extracellular matrix (ECM) supports the organ
 - Contains blood vessels, lymphatics, nerves
 - **Tissues separated by basement membrane (BM)**
 - **Often a *lumen* (fluid- or air-filled cavity)**
 - Transports secretory products from/to epithelium
 - **stroma-BM-epithelium microstructure: designed to maximise surface area of epithelium-lumen interface**
- **Notable biophysics and transport:**
 - **BM is a physical barrier to cell motion between tissues**
 - **Oxygen, glucose, and growth factors can only reach epithelium by diffusion**

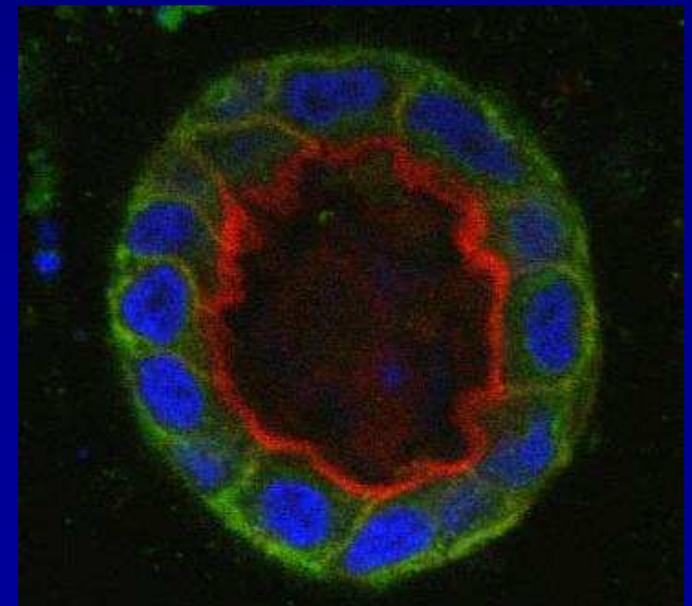
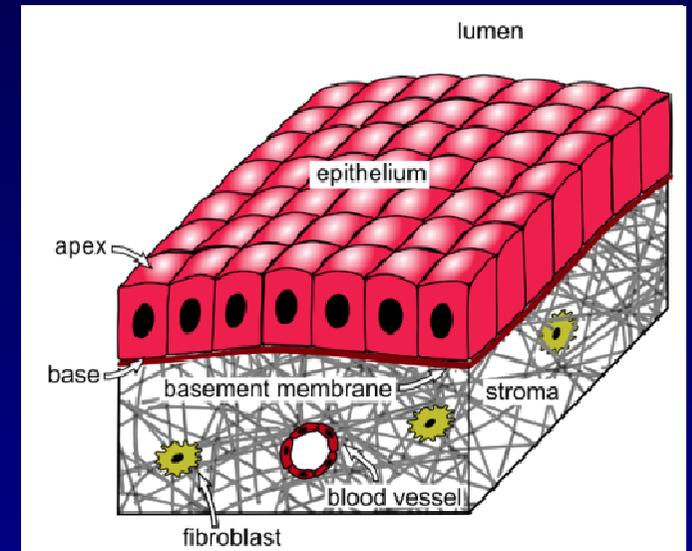


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- **Maintaining tissue structure**
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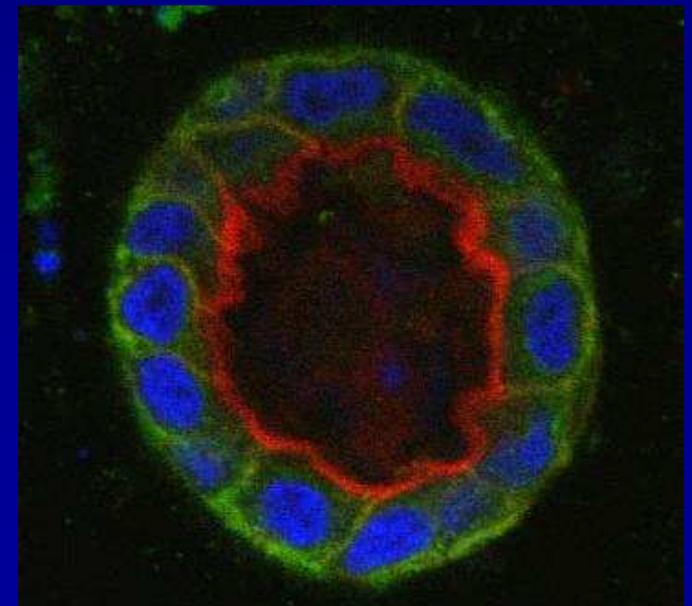
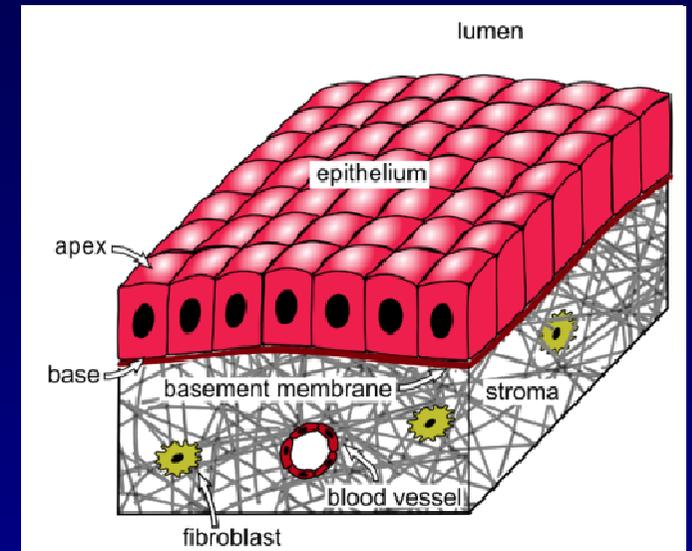
Maintaining Tissue Structure: Mechanics

- Mesenchymal cells (e.g., fibroblasts) can move freely in the stroma, secrete and degrade ECM
- Epithelial cells are polarised:
 - Anisotropic adhesion receptor distribution
 - Integrins on base for cell-BM adhesion
 - Heterophilic adhesion
 - No adhesion receptors on apex
 - E-cadherins on basolateral sides for cell-cell adhesion
 - Homophilic adhesion
- Mechanics determine tissue geometry:
 - Balance of cell-cell and cell-BM strength
 - Distribution of cell receptors likely help in determine curvature
 - Mechanics of cell-BM adhesion, stresses, BM-to-ECM coupling likely determines BM curvature.



Maintaining Tissue Structure: Population Dynamics

- Cell populations must be maintained in homeostasis
 - Proliferation to replace aged cells
 - Apoptosis to remove damaged cells, or those out of place
- Adhesion receptors are not merely mechanical:
 - E-cadherin helps detect presence/absence of neighbours
 - Trigger proliferation when missing a neighbour (E-cadherin/ β -catenin)
 - Suppress cell cycle when attached to neighbours (block Cyclin D1, etc.)
 - Integrins detect detachment from BM
 - Trigger apoptosis (anoikis)
 - (This is why they're receptors. 😊)

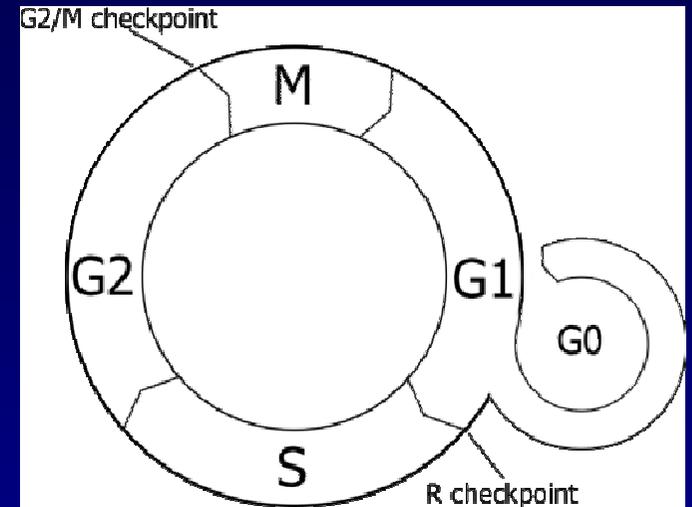


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Cell Birth and Death: Proliferation

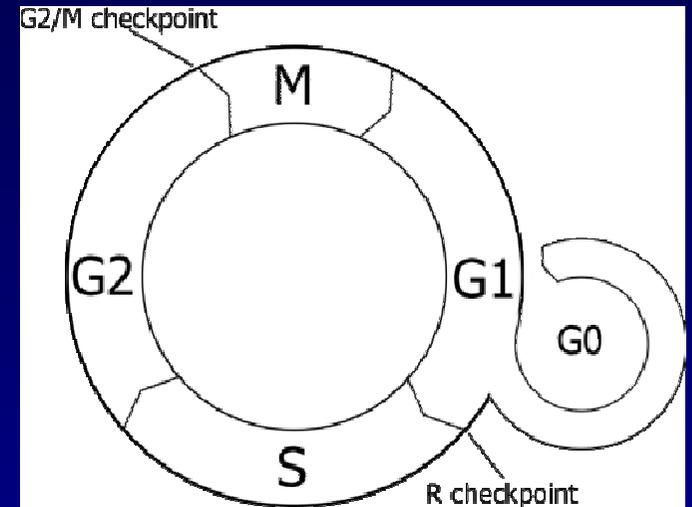
- **Tightly-controlled cell-cycle:**
 - **G₀:** quiescent “resting state”
 - **S:** Synthesis of new DNA
 - **G₂:** “gap” phase – final prep for division
 - **M:** mitosis phase
 - DNA divided into two daughter nuclei (mitosis)
 - Cytoplasm and organelles divided into daughter cells (cytokinesis)
 - **G₁:** “gap” or “growth” phase – daughter cells grow in volume, then exit cycle
 - **Note 1:** Some biologists treat G₀+G₁ as long, variable-length G₁ phase (often seen in flow cytometry)
 - **Note 2:** Others treat G₁ as relatively fixed length, with G₀ of variable length (useful for Ki-67 matching)



Cell Birth and Death: Proliferation

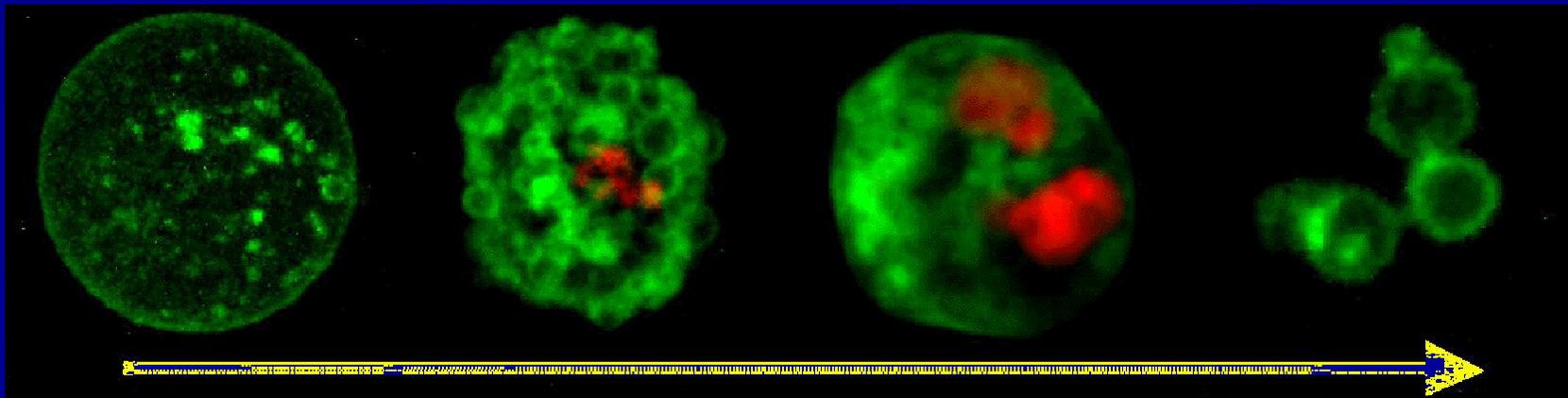
- **Checkpoints provide opportunities for:**
 - **Cycle arrest: Restriction checkpoint R at G1/S**
 - **DNA error checking and repair**
 - **Apoptosis for irreparable damage**

- **Cycle progression controlled by intracellular signalling**
 - **Cyclins and cyclin-dependent kinases (CDKs)**
 - **Connected to other signalling networks**



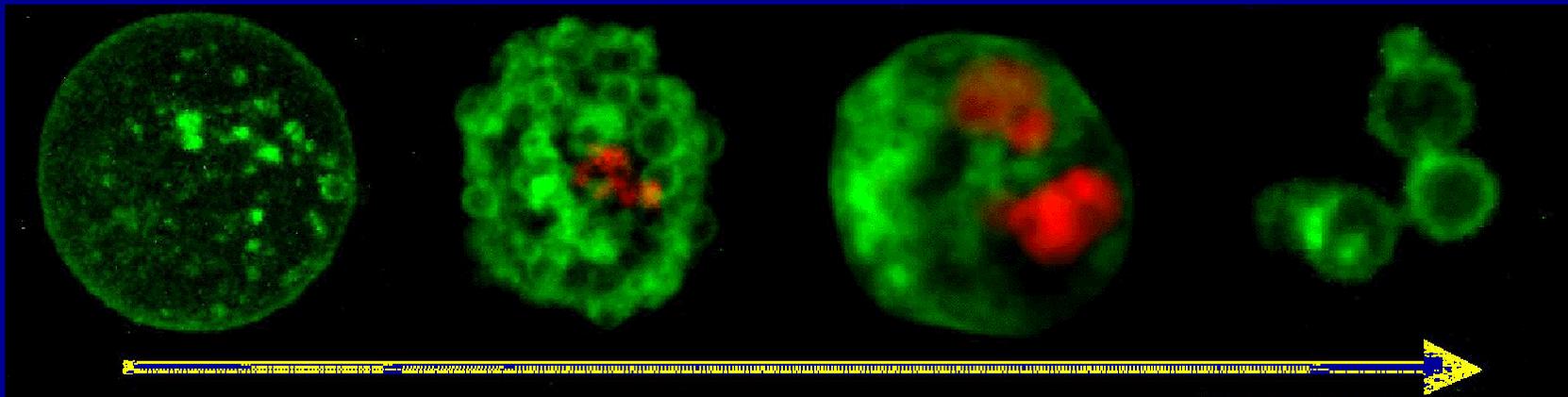
Cell Birth and Death: Apoptosis

- **Apoptosis: orderly cell death**
 - Self-regulated, in response to signalling events
 - Early processes:
 - Mitochondria lose membrane potential, integrity
 - Pre-positioned caspases are activated (cleaved), begin degrading cell
 - Orderly cell volume shrinking
 - requires active Na^+ , K^+ pumps!



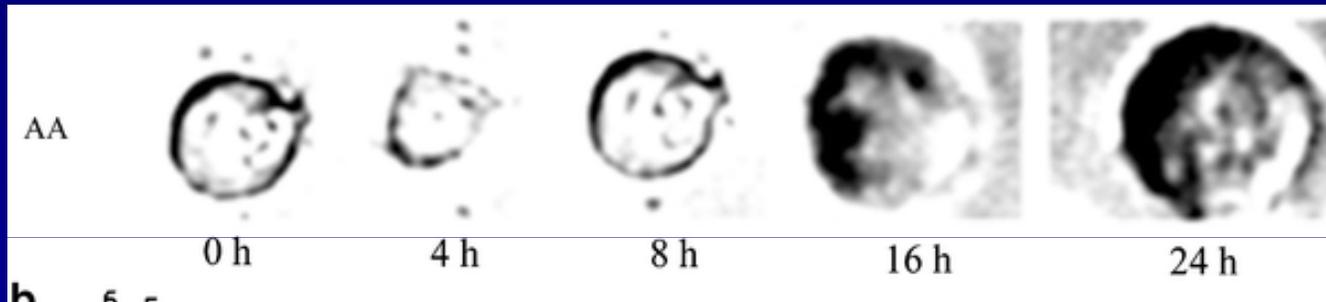
Cell Birth and Death: Apoptosis

- **Apoptosis: orderly cell death (continued)**
 - DNA is chopped into pieces
 - Organelles disassembled
 - Cell contents encapsulated into apoptotic bodies
 - Protects surrounding cells from otherwise damaging reactions
 - Cell lyses to release apoptotic bodies
 - Apoptotic bodies phagocytosed
 - Entire process requires energy!

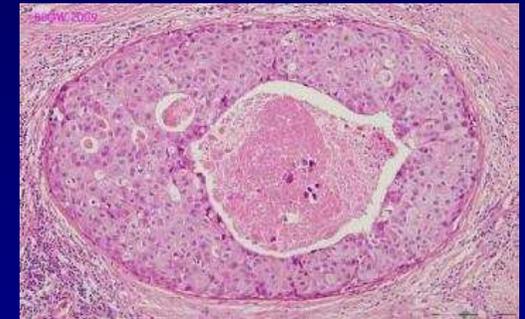


Cell Birth and Death: Necrosis and Calcification

- **Necrosis: uncontrolled cell death**
 - Can occur due to energy depletion (hypoxia or hypoglycemia), mechanical injury, chemical stressors, failure during apoptosis, etc.
 - Uncontrolled cell volume
 - No energy for Na^+ , K^+ pumps
 - Cell swells, then bursts
 - Cell contents released into microenvironment



- No orderly disassembly of nucleus, organelles, etc.
- Generally no phagocytosis
- **Calcification:**
 - Ca^+ pumps not active
 - Solid cell fraction replaced by calcium phosphate and oxalate crystals
 - Hard microcalcifications results



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Cell Signalling

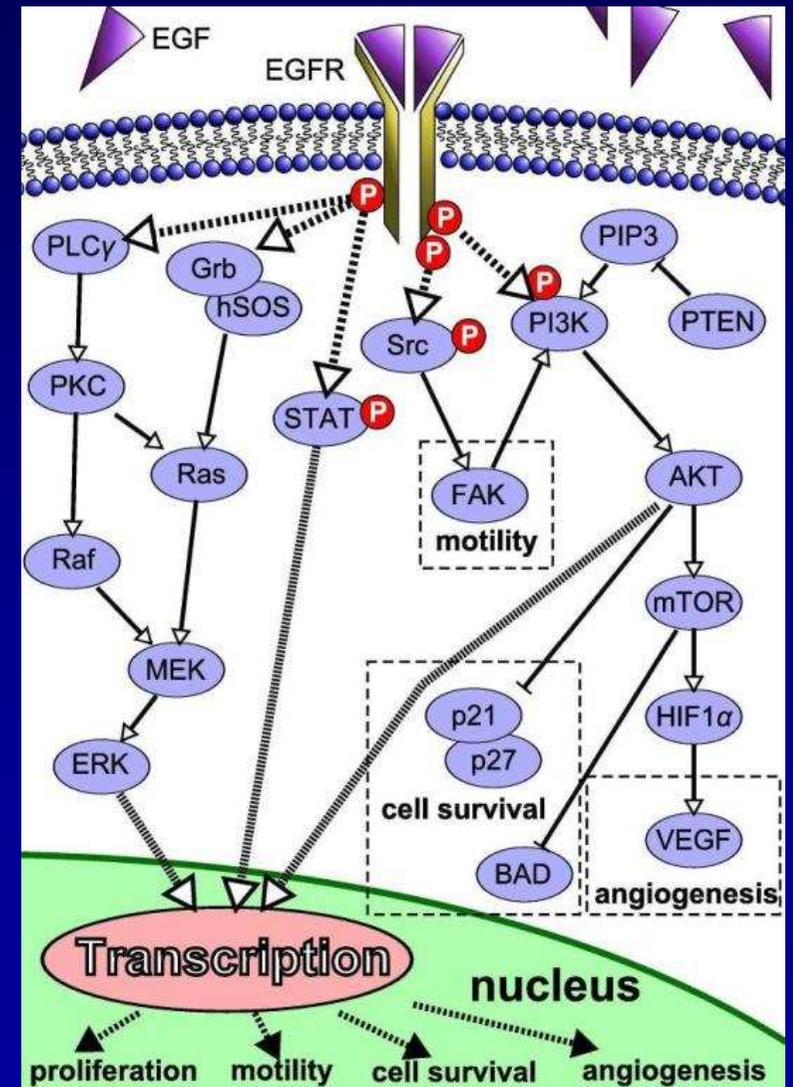
- Cells are regulated by internal signalling networks
 - DNA transcribes RNA
 - RNA encodes proteins
 - Proteins:
 - Assemble into structures
 - Perform “duties”
 - Transmit information through reactions
 - Network:
 - Redundancies
 - Feedback loops
 - Signal amplification

Cell Signalling: Two examples

- HIF signalling:
 - Cells create hypoxia-inducible factors (HIFs)
 - Big example: HIF-1 α
 - Ordinarily, O₂ tags these for degradation
 - During hypoxia, HIFs accumulate → HIFs as O₂ sensors
 - Trigger downstream transcription
 - Decreased cell adhesion
 - Increased motility
 - Glycolysis
 - Temporary resistance to apoptosis
 - VEGF secretion

Cell Signalling: Two examples

- EGFR signalling
 - Epidermal growth factor (EGF) binds to EGFR receptor
 - Ligated EGFR receptors dimerise
 - Dimerised EGFR receptors phosphorylate intracellular proteins
 - Downstream actions:
 - Transcription
 - Increased motility
 - Increased proliferation
 - Cell survival

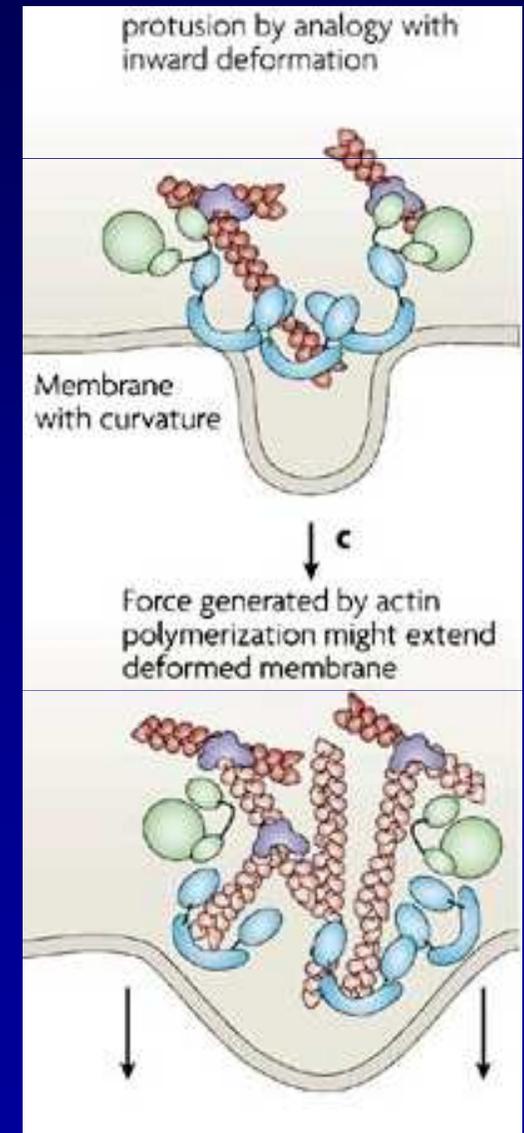
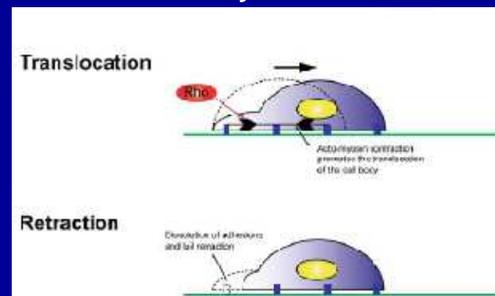
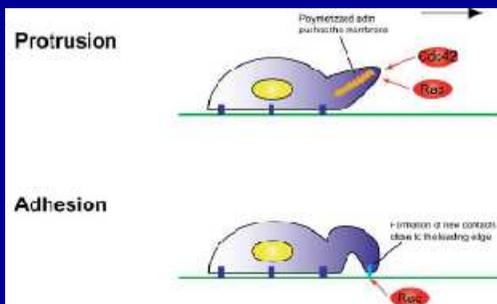


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Cell Motility

- Actin polymerisation and depolymerisation:
 - Actin monomers join or leave chains to grow or shrink the cytoskeleton
- Cell signalling can nucleate biased polymerisation (e.g., in response to a gradient)
 - EGFR → Src → N-Wasp → Arp2/3 nucleation → ...
- Actin fibres deform and extend cell membrane, forming a pseudopod (“false foot”)
 - Filopodium: finger-like projection
 - Lamellipodium: sheet-like projection
- Cells focally secrete MMPs from invadopodia:
 - Degrade ECM
 - Break integrin bonds
 - Create space for motion
- Cell forms new focal adhesion on leading edge
- Actin polymers broken down in trailing edge (depolymerisation)
- Cell contracts to pull towards leading edge
- Protrusion – Adhesion – Contraction – Retraction cycles



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Oncogenes and Tumour Suppressor Genes

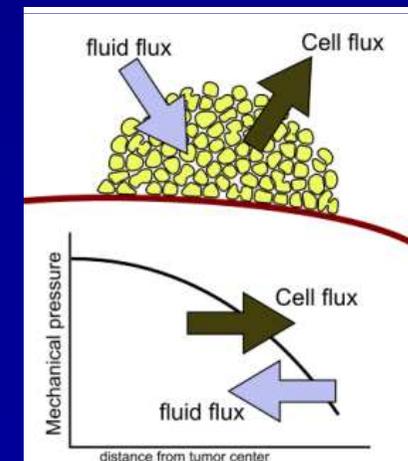
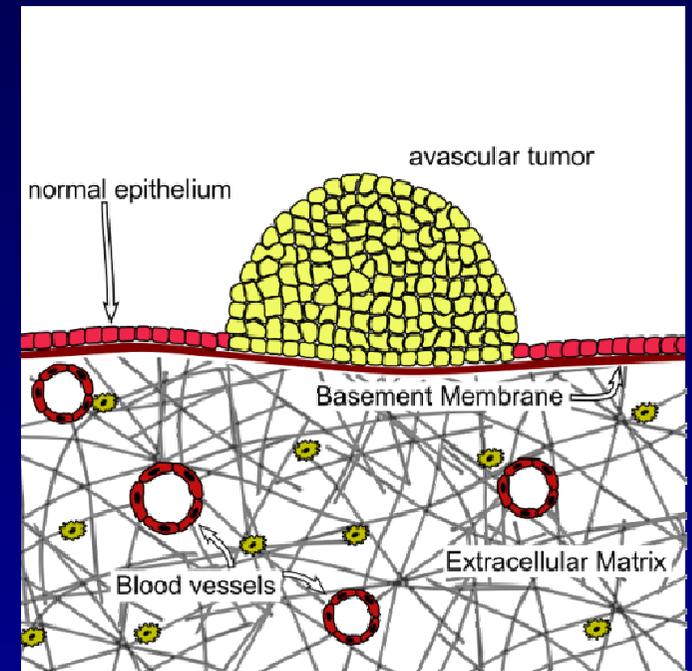
- Oncogenes:
 - Proliferation-promoting genes : “gas pedal”
 - Examples:
 - Growth factor secretion (including autocrine): EGF, PDGF, VEGF
 - Internal promotion of cell cycle: β -catenin promotes transcription of Cyclin D1
 - Counter-act TSGs: MDM2 degrades p53
 - Growth factor receptors: EGFR / ErbB
 - Downstream regulators of receptor pathways: Ras, Raf, Src
- Tumour suppressor genes (TSGs):
 - Impede proliferation : “brake pedal”
 - Examples:
 - Block oncogenic signals: VHL helps degrade HIF-1 α
 - Regulate cell cycle / create inhibitory signals: Rb impedes cell cycle, promotes arrest at G1/S
 - Repair DNA damage: TP53 impedes cell cycle to allow DNA repair at G1/S
 - Promote apoptosis: p53 can trigger apoptosis
 - Knudson 2-hit hypothesis:
 - You have 2 copies of each TSG, so need to eliminate both copies to lose function
 - Discovered in studying Rb (retinoblastoma) TSG
 - Mitigating factor: loss of heterozygosity
 - Mitigating factor: partial loss of function

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Abusing the system: Carcinogenesis

- **Mutations and/or epigenetic events:**
 - **Down-regulate apoptosis (lose TSGs)**
 - No anoikis – cells can survive in the lumen
 - p53 mutation – cells can ignore apoptosis signals
 - **Up-regulate proliferation (oncogenes)**
 - Decreased contact inhibition – proliferate even in the presence of neighbours
 - Increased secretion of growth signals → self-signalling
 - “stuck switches”
 - HER2 – can dimerise without binding ligand
 - » increases signalling activity
 - K-ras mutation – constitutive active, allowing EGFR signalling with EGF
- **Consequences:**
 - Increased survival fitness versus normal cells
 - Forms a colony of hyperproliferative cells
- **Mechanics:**
 - Mechanical pressure, stresses created by proliferating cells
 - Net outward flux of cells as mass grows – Darcy’s law
 - Net inward fluid flux



Abusing the system: Avascular Growth

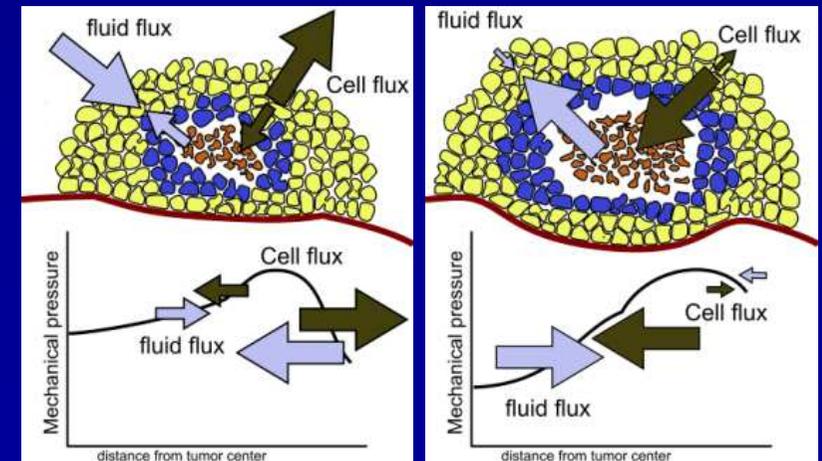
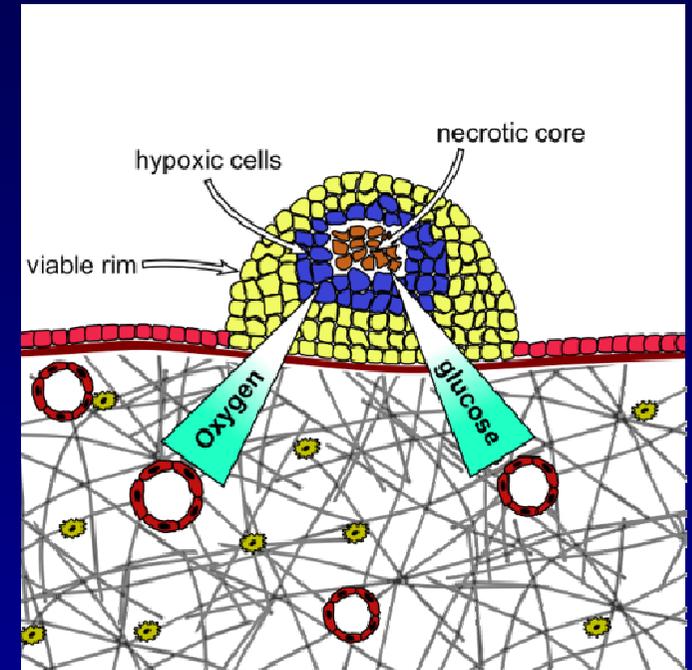
- Can only receive substrates by diffusion from the stroma
 - Substrate gradients
 - Hypoxia
 - Hyperglycemia

- Consequences:

- Viable rim thickness related to diffusion length scale
- Heterogeneous growth rates – relationship with oxygen and glucose (cell energetics)
- Inner band of hypoxic cells
 - HIF signalling
- Interior necrotic core

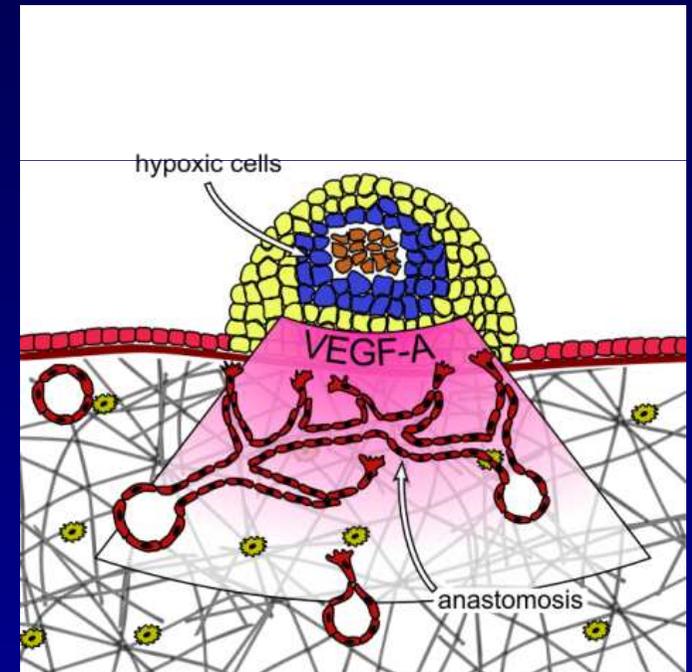
- Mechanics:

- Outward fluid flux from necrotic core due to lysis
- Inward cell flux due to reduced interior strain
- Steady tumour volume:
 - Cell flux out of viable rim \approx fluid flux from necrotic core



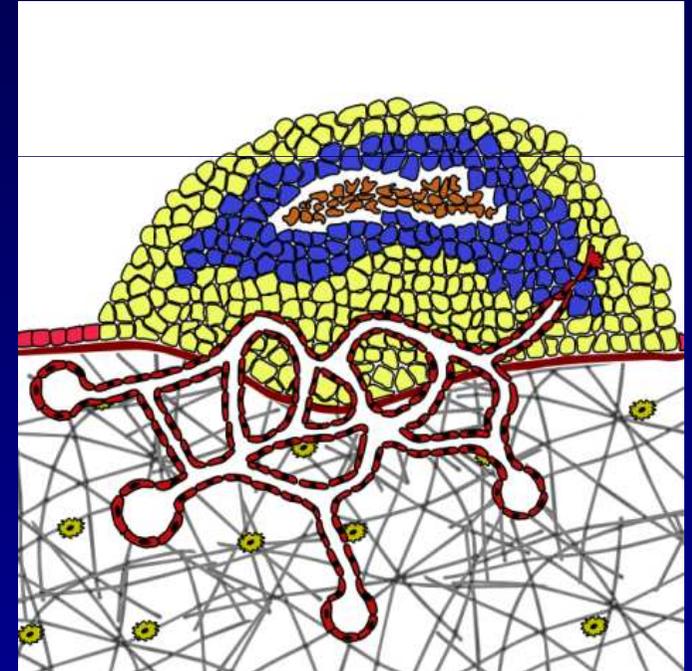
Abusing the system: Angiogenesis

- Hypoxic cells release angiogenic factors
- VEGF diffuses into stroma
- Endothelial cells respond to VEGF
 - Degrade vessel walls
 - Chemotaxis (up ∇ VEGF), haptotaxis (up ∇ ECM)
 - Increased proliferation
 - Temporary suspension of anoikis (to facilitate survival until new basal lamina surrounds mature vessels)
- New vessels grow towards tumour, generally towards the VEGF gradient
- New vessels cross-link (anastomose)
- New blood flow in the vessels – new substrate transport to fuel further tumour growth
- Vessels mature: pericytes recruited, deposit basal lamina around vessels



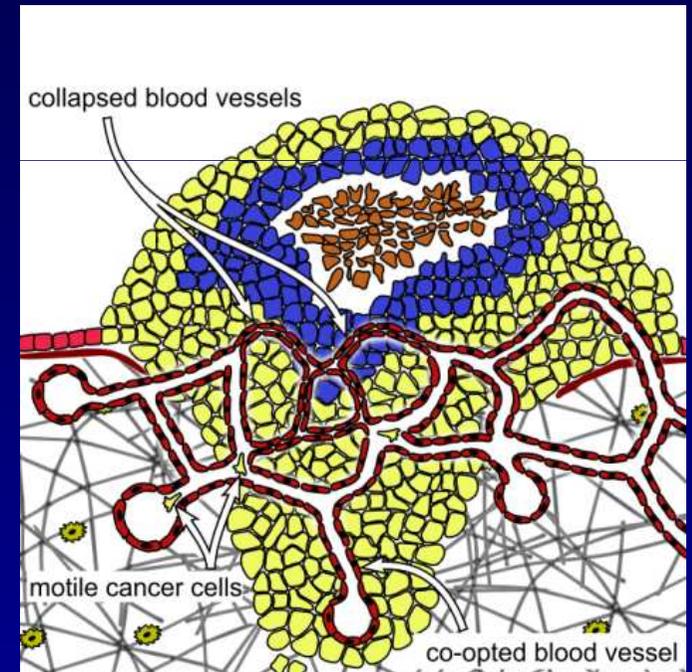
Abusing the system: Vascular Growth & Invasion

- **New vessels fuel rapid tumour growth**
- **BM deformation and stress**
- **Hypoxic stress remains**
 - Glycolysis
 - Acidosis
 - *Selection pressures*



Abusing the system: Vascular Growth & Invasion

- **Mutations**
 - Acid-resistant
 - BM and ECM degradation (by MMPs)
 - Motility
- **Invasion into stroma**
 - By growth and by motility
 - Co-option of blood vessels
- **Sustained tissue stress**
 - Collapsed blood vessels
 - New rounds of angiogenesis
- **Metastasis**
 - Travel through blood vessels and lymphatics
 - Extravasation, growth of new tumour
 - Possible role of metastatic niche



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Coming Next:

- **Lecture 1:**
 - Cancer biology for modellers
- **Lecture 2:**
 - **An agent-based cell model; application to DCIS**
- **Lecture 3:**
 - Parameter estimation, patient-specific calibration
- **Lecture 4:**
 - Numerical method, simulation results

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Some References

- Some biology texts:
 - P. Macklin. Biological background. In: V. Cristini and J. Lowengrub. *Multiscale Modeling of Cancer*. Cambridge University Press, Cambridge, UK, 2010. Chapter 2, pages 8-24. ISBN 978-0521884426. (in press)
 - B. Alberts et al. *Molecular Biology of the Cell*. Garland Science, New York, NY USA, 5th edition, 2007. ISBN 978-0815341116.
 - M. Knowles and P. Selby, eds., *Introduction to the Cellular and Molecular Biology of Cancer*. Oxford Univ. Press, Oxford, UK, 4th edition, 2005. ISBN 0-19-852563-X.
- Also see some great modelling texts by Wodarz & Komarova, Anderson et al.
- Some great websites:
 - Cell biology and animations: <http://www.johnkyrk.com/>
 - SIU histology: <http://www.siumed.edu/~dking2/>
 - Zoomified histology:
<http://www.meddean.luc.edu/lumen/MedEd/Histo/virtualhistology.htm>

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- (new but under construction)

- <http://biomathematics.shis.uth.tmc.edu>

- (old but already built)